User Guide
Standard Quantitative Cartridge Kit (C105201-Q)

A. Specifications

Specification	Description
DNA Sizing Range	20-5,000 bp
DNA Sizing Precision*	2% CV
DNA Quantitative Precision*	4% CV
Sample Number	200 runs
Shelf Life	6 months

* Precision is determined by the 15-622 DNA Size Marker (C109200).

B. Kit Components and Storage Conditions

Item	Storage Condition
Standard Quantitative Cartridge (C105201-Q)	4-30°C (Do Not Freeze)
20-1,500 bp Quantitative Marker (C109109-500Q, 500 µL)	Short-Term (≤ 3 months): 4-30°C Long-Term (> 3 months): -20°C
15-622 bp Size Marker (C109200-100, 100 µL)	Short-Term (≤ 3 months): 4-30°C Long-Term (> 3 months): -20°C
Separation Buffer (C104406, 50 mL)	4-30°C
Dilution Buffer (C104405, 15 mL)	4-30°C
Mineral Oil (C104404, 8 mL)	4-30°C

⚠ **Please always store cartridges in a light-proof bag, and then store in the cartridge box after analysis.**

⚠ **To maintain marker stability, avoid repeated freeze-thaw cycles.**

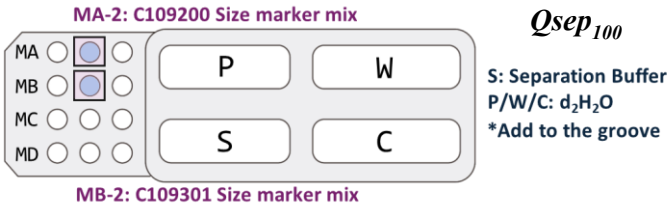
C. Cartridge Unpacking Preparation

The new cartridge must undergo **HV check** and **calibration**. Please follow the instructions provided in the unpacking guide and calibrate using C109109 Quantitative Marker.

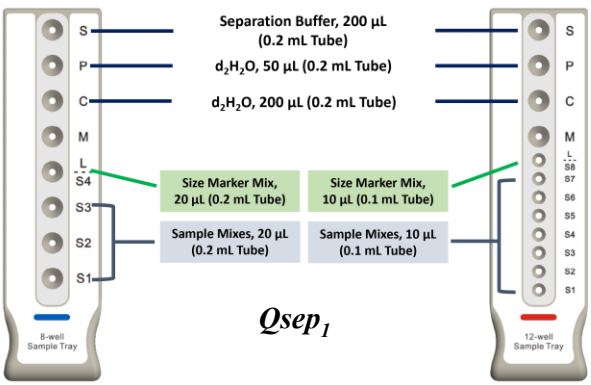
- Calibration mix for cartridge unpacking:
  - 20 bp-1,500 bp Quantitative Marker (C109109): 5 µL
  - Dilution Buffer (C104405): 15 µL

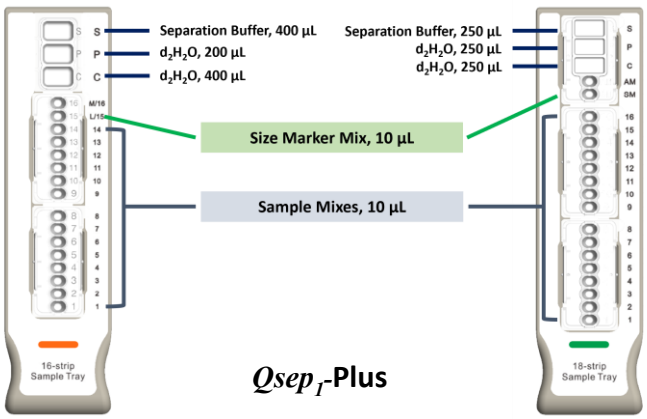
⚠ **CAUTION:** To prevent cartridge lockout after 10 failed calibration attempts, please contact your local distributor or BiOptic if the issue persists after following the troubleshooting steps in the cartridge unpacking guide.

D. Buffer, Marker, and Sample Preparation



⚠ **Hold the holder steady and press the alignment marker tube firmly into the well with your thumb.**





⚠ **Ensure the buffer tray is pushed to the end until the color bar aligns with the edge of the holder.**

1. Compatible Sample Tubes

	Name	Cat. No.	Volume	Image
Qsep100 / 8-well Sample Tray	0.1 mL PCR Tube	-	20 µL	
	0.2 mL PCR Tube	-		
12-well Sample Tray	0.1 mL Strip Tube	C104252		
16-strip Sample Tray	16-strip Sample Tube	C104254		
18-strip Sample Tray	18-strip Sample Tube	C104257		

2. Size Marker Mix for Different Size Ranges

- For Sample Size Range from 20 bp to 1,500 bp:
  - 20 bp-1,500 bp Quantitative Marker (C109109): 5 µL
  - 15-622 bp Size Marker (C109200): 10 µL
  - Dilution Buffer (C104405): 5 µL
- For Sample Size Range from 20 bp to 5,000 bp:
  - 20 bp-5,000 bp Quantitative Marker (C109102): 5 µL
  - 50 bp-3,000 bp Size Marker (C109301): 10 µL
  - Dilution Buffer (C104405): 5 µL

3. Sample Mix Preparation

- Quantitative Marker (C109109/C109102): 5 µL
- Sample: X (2～15) µL
- Dilution Buffer (C104405): 15-X (0～13) µL

4. Recommended Sample Concentration

Fragmented sample: 0.2-50 ng/µL [Best: 0.5~5 ng/µL]

Smear sample: 1-50 ng/µL [Best: 2~20 ng/µL]

ℹ The recommended sample concentration indicates the final concentration for loading onto the instrument.

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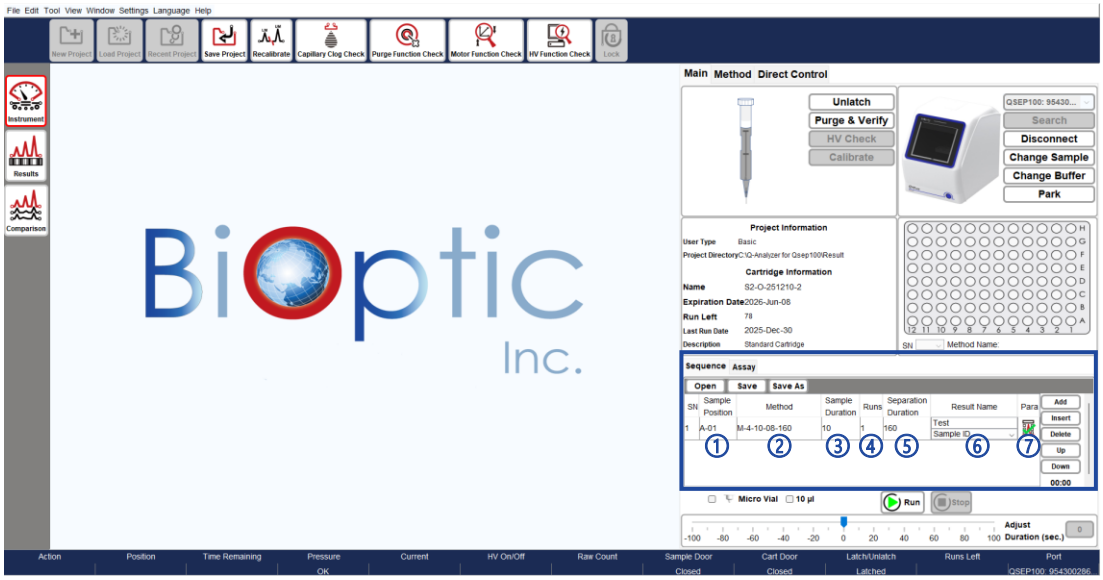
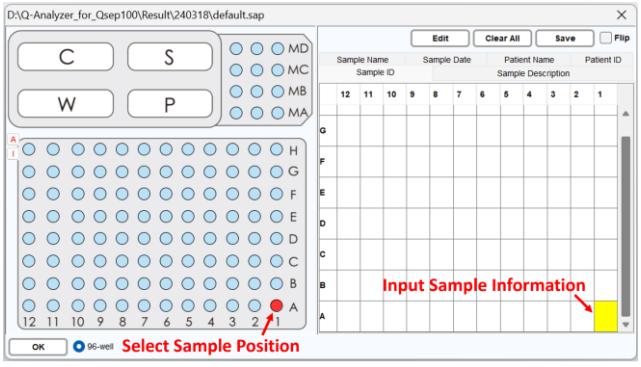
I. Software Operation Guide

Qsep100



① Sample Position: Place the sample and select the corresponding position. Input sample information if necessary.

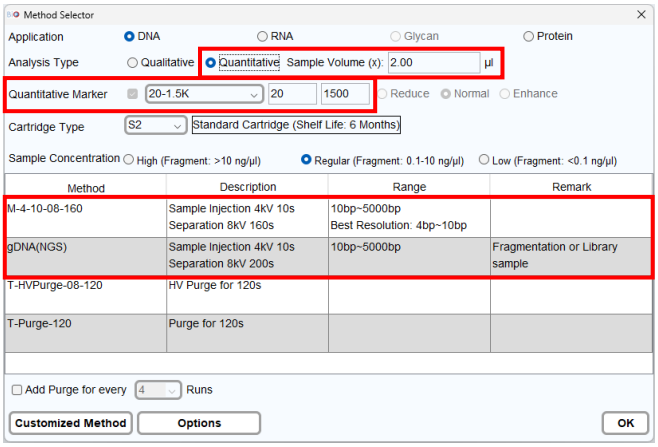


② Method:

(a) Analysis Type to **Quantitative** and input the sample volume for the sample mix.

(b) Select Quantitative Marker.

(c) Select analytic Method in the Method Selector.



- **Adjust injection conditions based on sample concentration.**

Sample Concentration	High (2kV, 10s)	Regular (4kV, 10s)	Low (8kV, 10s)
Fragmented DNA	> 10 ng/µL	0.1-10 ng/µL	0.01-0.1 ng/µL
Smear DNA	> 50 ng/µL	2-50 ng/µL	0.5-2 ng/µL

③ Sample Duration: Adjust the sample injection time to increase/decrease injection amount.

- **Modify injection conditions based on sample concentration.**

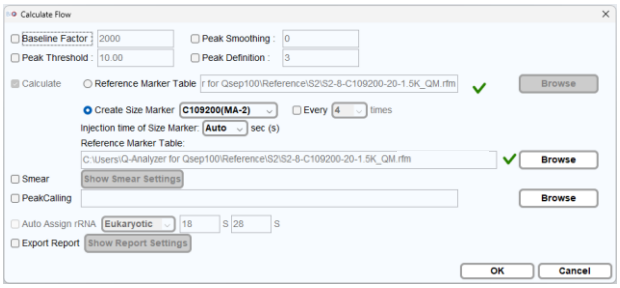
④ Runs: Set the repetition time.

⑤ Separation Duration: Adjust the duration to extend/reduce the separation time.

(Optional)

⑥ Result Name: Input the result name for the result file.

⑦ Para: Choose between (a) Reference Marker Table and (b) Create Size Marker for calculation.



- **When using "Create Size Marker" function, select the appropriate size marker you use. For example, "20-1.5k" is paired with C109200, "20-5k" is paired with C109301.**

8. Click "Run" to start the sequence analysis.

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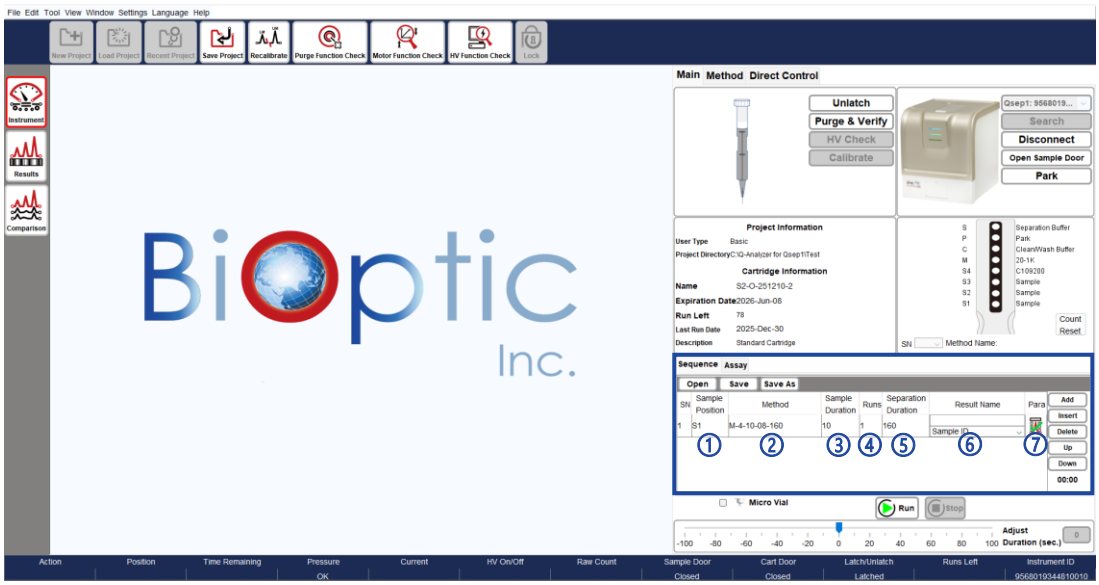
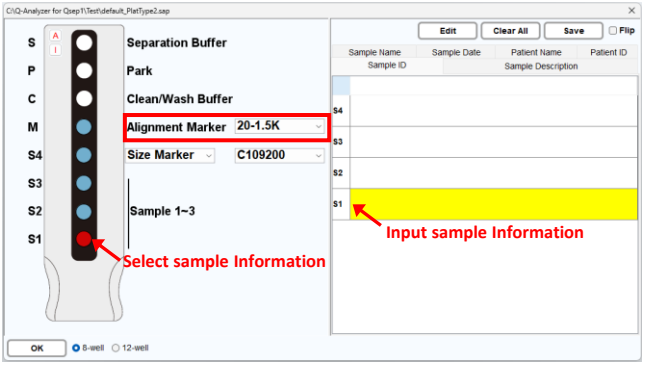
Qsep$_1$



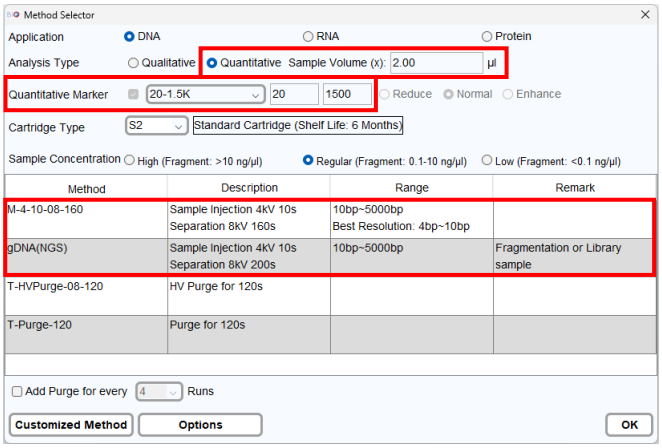
- Click “Open Sample Door” and insert the desired sample tray
- Click “Park” to move the sample tray into position.

Qsep1	*Qsep1*-Plus
8-well Sample Tray 12-well Sample Tray	8-well Sample Tray 12-well Sample Tray 18-strip Sample Tray

① Sample Position:
- Choose Alignment Marker type (20-1.5K or 20-5K).
- Place the paired Size Marker Mix at S4 position if required, and input the sample information if necessary.



② Method:
(a) Analysis Type to **Quantitative** and input the sample volume for the sample mix.
(b) Check Quantitative Marker.
(c) Select analytic Method in the Method Selector.



- **Adjust injection conditions based on sample concentration.**

Sample Concentration	High (2kV, 10s)	Regular (4kV, 10s)	Low (8kV, 10s)
Fragmented DNA	> 10 ng/µL	0.1-10 ng/µL	0.01-0.1 ng/µL
Smear DNA	> 50 ng/µL	2-50 ng/µL	0.5-2 ng/µL

③ Sample Duration: Adjust the sample injection time to increase/decrease injection amount.
- **Modify injection conditions based on sample concentration.**

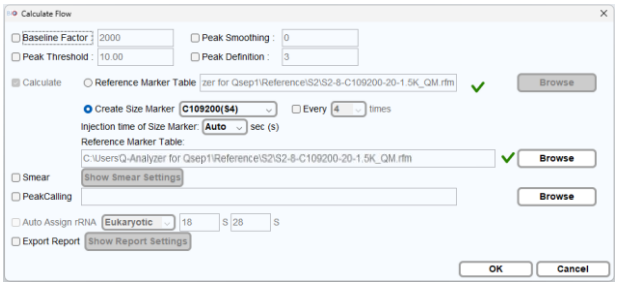
④ Runs: Set the repetition time.

⑤ Separation Duration: Adjust the duration to extend/reduce the separation time.

(Optional)

⑥ Result Name: Input the result name for the result file.

⑦ Para: Choose between (a) Reference Marker Table and (b) Create Size Marker for calculation.



- **When using “Create Size Marker” function, select the appropriate size marker you use. For example, “20-1.5k” is paired with C109200, “20-5k” is paired with C109301.**

8. Click “Run” to start the sequence analysis.

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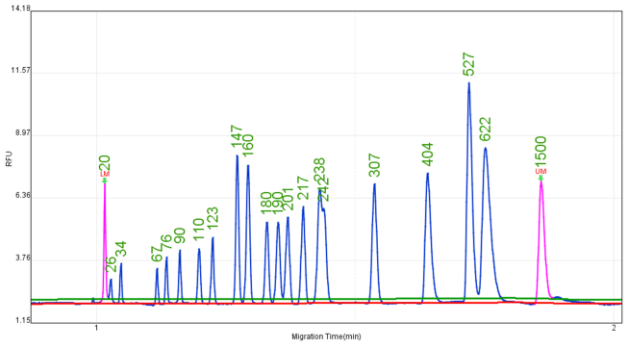
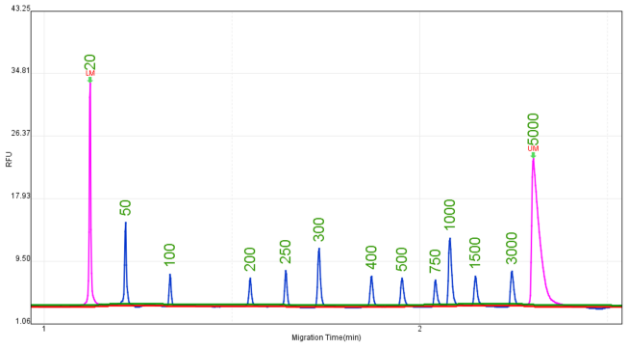
F. Result & Application

- **Quantitative Marker & Size Marker Result**

20-1.5K & C109200 15-622 bp Size Marker

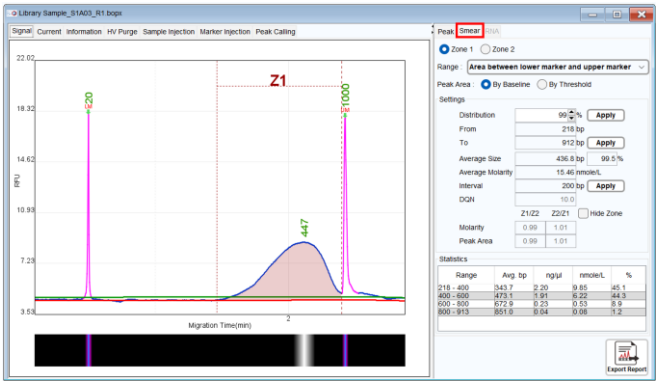


20-5K & C109301 50-3000 bp Size Marker



- **Fragmented & Library Sample Validation**

Use the smear function to obtain average size, molarity, and size distribution of fragmented DNA or library DNA samples.



Distribution:	99.5 %	**Average Size:**	436.8 bp
From:	218 bp	**Average Molarity:**	15.46 nmole/L
To:	913 bp	**DQN:**	10.0

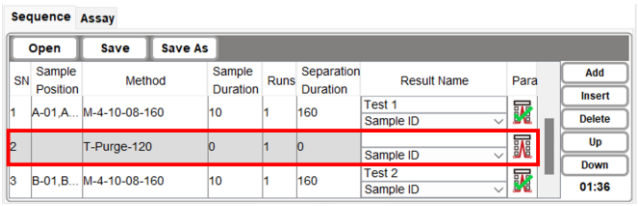
Range	Avg. bp	ng/µl	nMole / L	Percent
218 - 400	343.7	2.20	9.85	45.1
400 - 600	473.1	1.91	6.22	44.3
600 - 800	672.9	0.23	0.53	8.9
800 - 913	851.0	0.04	0.08	1.2

G. Troubleshooting

Before attempting any troubleshooting, ensure that the system is functioning properly and that all operations are following the instructions.

If encountering unstable current during sample injection or separation steps, which may be caused by unknown substances in PCR reagent buffer or other kit buffers, consider the following solutions:

1. Dilute the sample using dilution buffer.
2. Centrifuge the sample for a period to allow residues to accumulate at the bottom of the tube.
3. Insert a "T-purge-120" method between several sample runs. For example, insert one run of "T-Purge-120" every 5-10 sample runs.



H. Cartridge Disposal

Please wear gloves before discarding the cartridge.

1. Bend the cartridge tip.
2. Open the cap on the gel reservoir and remove the inner cap.
3. Pour the gel into the chemical waste container.
4. Dispose of the cartridge in the trash bin.

Contact Information:
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