User Guide

Qexp-MDx NQO1 Lyophilized Genotyping Kit

Cat. C9051101-024-O

Research Use Only

Product Information

Contents and Storage

Table 1 NQO1 Lyophilized Kit

Label	Component	Amount	Storage
		24 rxns	
Specimen Collection			
	Oral Swab with Collection Tube contains 3mL UTM[a]	24 Pcs	RT
Direct Lysis Assay			
PK	Proteinase K	1 Pc	-20℃
L2	DNA Lysis Buffer 2.0	4800 µL	-20℃
Multiplex PCR Assay			
	Direct PCR Lyo-PreMix-NQO1	3 Pcs (8rxns/Pc)	-20℃
PC	Positive Control-NQO1	300 µL	-20℃
W	Nuclease-Free Water	400 µL	-20℃

[a]Oral Swab with Collection Tube containing 3mL UTM (C9060001-024) should be purchased separately.

- All components will be shipped at ambient temperature. For long-term storage, they should be stored at -20℃.

Guidelines for sample collection-Oral Swab

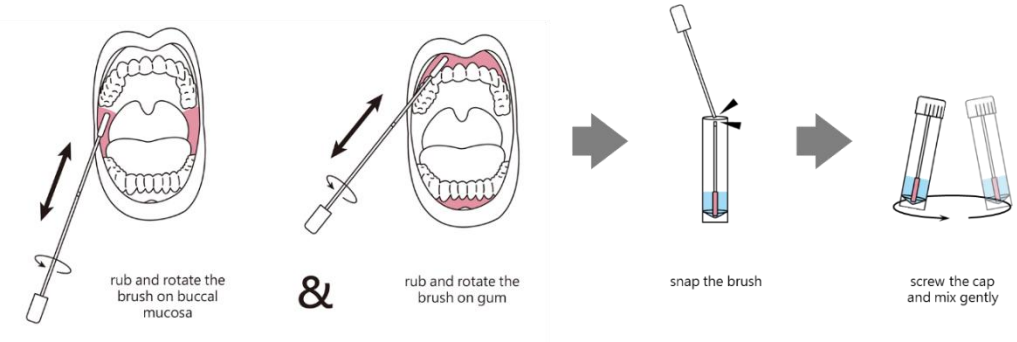
IMPORTANT

- Avoid drinking milk or having food rich in calcium before oral swab collection.
- Before sample collection, rinse the mouth with clean water for 10 seconds to remove microorganisms, food and drink remnants.
- Ensure that there is no eating, drinking, smoking, chewing gum, brushing teeth, or use of mouthwash after rinsing the mouth.

Oral specimen collection from the subject

1. Open the Specimen Collection Brush (Swab).
2. Force the brush a little bit on your buccal mucosa, then rub and rotate the brush on the buccal mucosa and gum 10 times on each side to collect cells.
3. Insert the brush into the supplied Collection Tube with UTM.
4. Snap the brush in the tube, then screw the cap of the Collection Tube.
5. Mix well gently to release the cells on the brush into UTM.

※ Can be stored at -20℃ for a week.



Perform Assay using $Qsep_{Ultra}$™ Otsuka

For the Installation, System Setup, Operation and Cartridge Verification of $Qsep_{Ultra}$, please refer to the operation manual (F0137_$Qsep_{Ultra}$ Otsuka User Manual) and pay attention to the safety information before use.

Instrument and Software

Table 2 Instrument and software compatibility

Instrument	Q-Analyzer™ for $Qsep_{Ultra}$
$Qsep_{Ultra}$™ Otsuka (C500001-O)	Version 1.1.1 or later

Proteinase K reconstitution

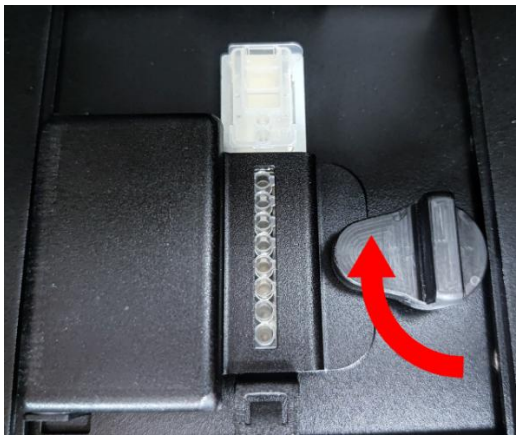
1. Spin down the Proteinase K and Nuclease-Free Water briefly.
2. Resuspend the lyophilized Proteinase K cake with 100 µL of Nuclease-Free Water.
3. Gently pipette the mixture and spin down the Proteinase K again briefly.

※ Stored at -20℃ after resonstitution.

Prepare oral specimen lysate

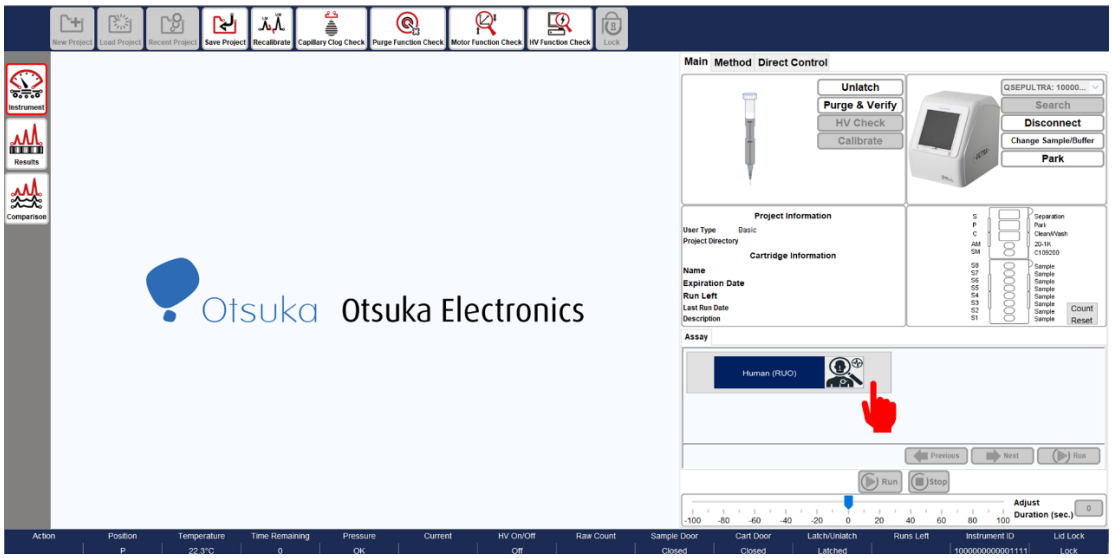
1. Gently invert the Collection Tube five times to ensure it mixes well.
2. Take 20 µL mixture from the Collection Tube into a clean PCR tube.
3. Add 2.6 µL Proteinase K, mix well, and incubate at room temperature for 1 min.
4. Set up samples in the well of 0.2 mL PCR tube (marked in red frame as follows).

5. Close the lid and twist the lid lock clockwise to ensure the lid is fixed.

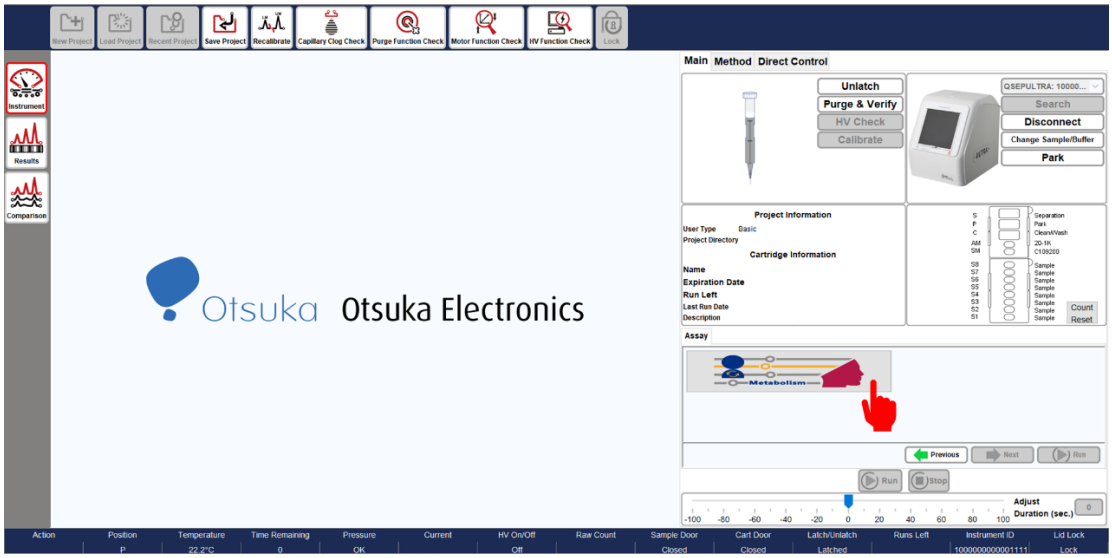


6. Close Sample Door and return to the software.

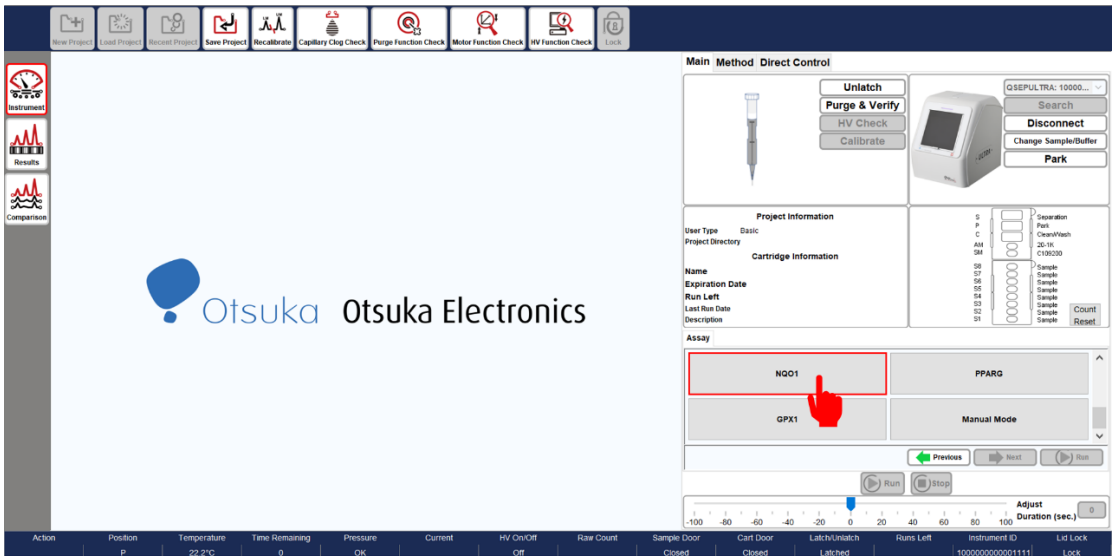
7. Click the icon **"Human (RUO)"** .



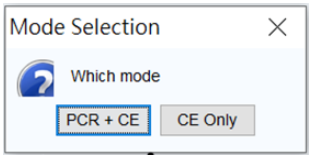
8. Click the icon **"Metabolism"** .



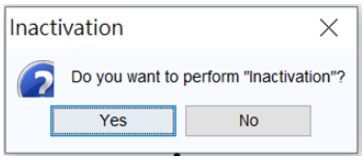
9. Click the icon **"NQO1"** to perform the assay.



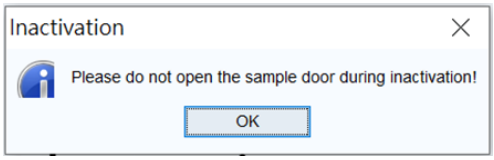
10. Click the **"PCR + CE"** mode.



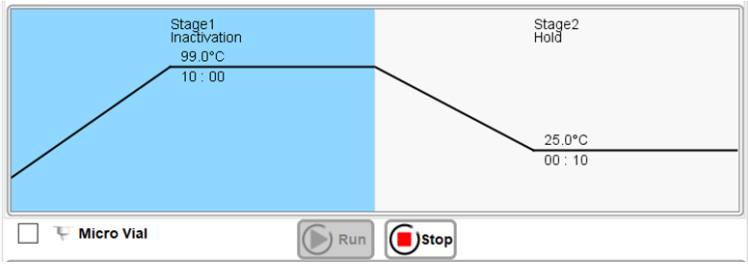
11. Click **"Yes"** to perform the inactivation step.



12. The system will mention **"Please do not open the sample door during inactivation!"**. Click **"OK"** to start the Proteinase K inactivation.



13. Waiting for the heating process to complete. If there is any emergency while heating, please click **"Stop"** (the button with a red square) to shut down the program.



14. The system will **"beep"** twice when the inactivation step is completed. And the tube holder will move to the door automatically.
15. Open the lid and take the inactivated samples out.
16. **Add 180 µL DNA Lysis Buffer 2.0** and mix well by gently vortexing or pipetting.
17. Incubate at room temperature for 10 minutes.

Prepare Direct PCR reaction

1. Reconstitute the lyophilized cake as the following table:

   Table 3 The preparation list of all PCR reactions

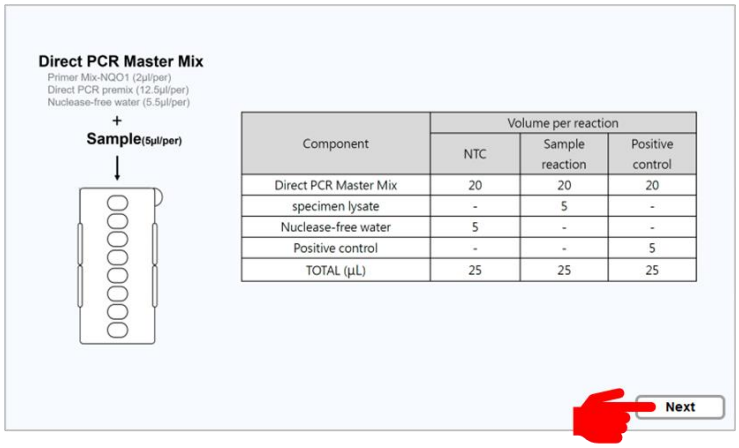
Component	Volume per reaction		
	NTC	Sample reaction	Positive control
Direct PCR Lyo-PreMix-NQO1	1 tube	1 tube	1 tube
specimen lysate	-	25	-
Nuclease-Free Water	25	-	-
Positive Control–NQO1	-	-	25
TOTAL (µL)	25	25	25

   ※ Spin down the Lyo-PreMix first before opening.

   ※ Do not use the same pipette to add water, specimen lysates, and positive control.

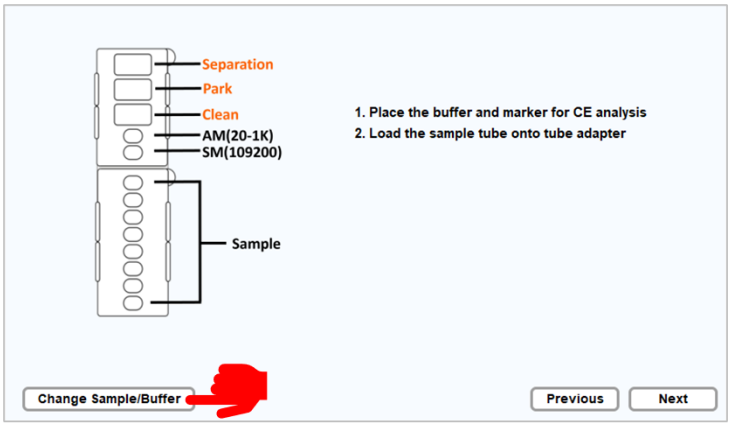
   ※ Always prepare the no template control (NTC) first, and the positive control last to avoid cross-contamination.

2. Pipetting and spin down (the lyophilisate will dissolve within seconds).
3. Transfer the samples to the $Qsep_{Ultra}$ 8-strip Sample Tube (C104258), then cover every tube with 10 µL mineral oil (provided with BiOptic's cartridge kit).
4. Briefly centrifuge the 8-strip Sample Tube to collect the liquid at the bottom of the tube.
5. Click **"Next"**.

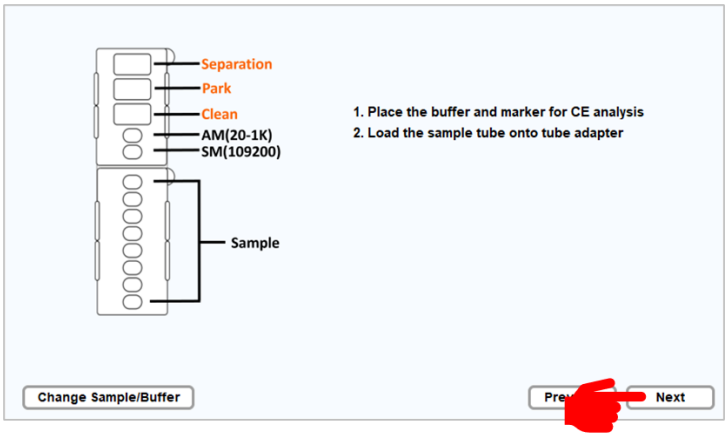


Setup and Run

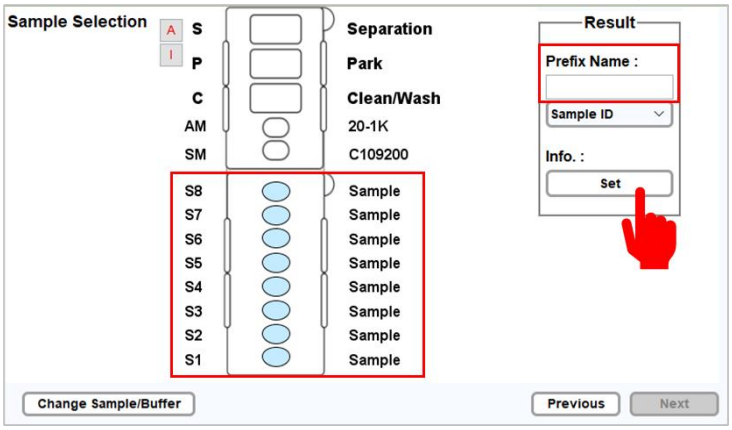
1. Click **"Change Sample/Buffer"** and place the prepared 8-strip Sample Tube (containing prepared 25 µL PCR reaction mix and covered with 10 µL mineral oil) on the tube holder.



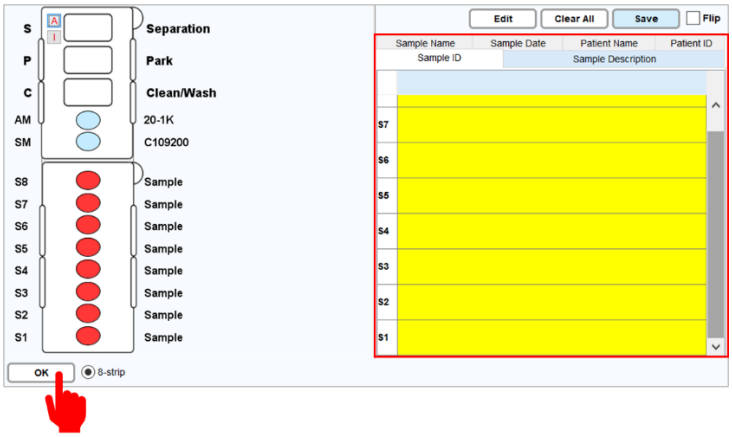
2. Click **"Next"**.



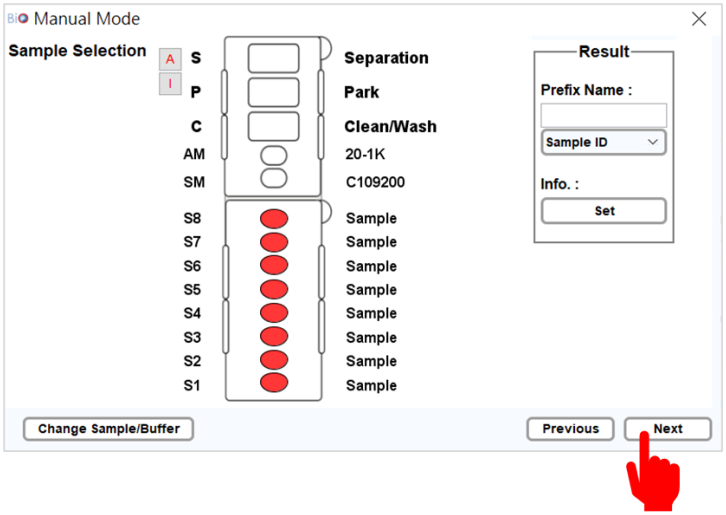
3. Select the sample position and input the result name in **"Prefix Name"** field. Then click **"Set"** to edit the information of the sample if necessary.



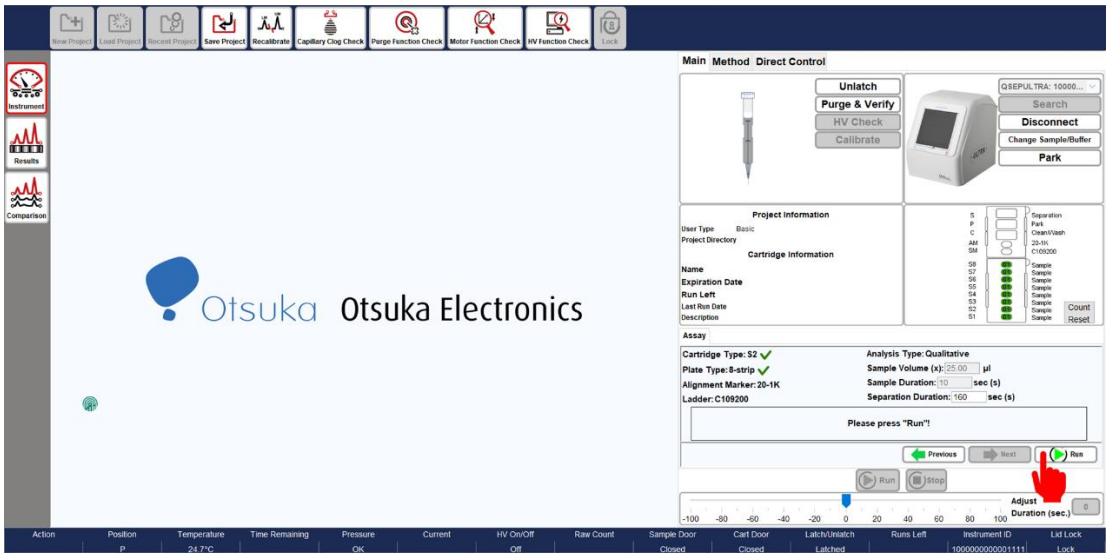
4. Select "**Sample Name**" or any options (**Sample Date**, **Patient Name**, etc.) you would like to record, then fill in the field and click "**OK**".



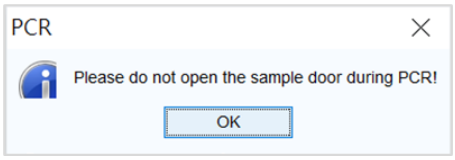
5. Click "**Next**".



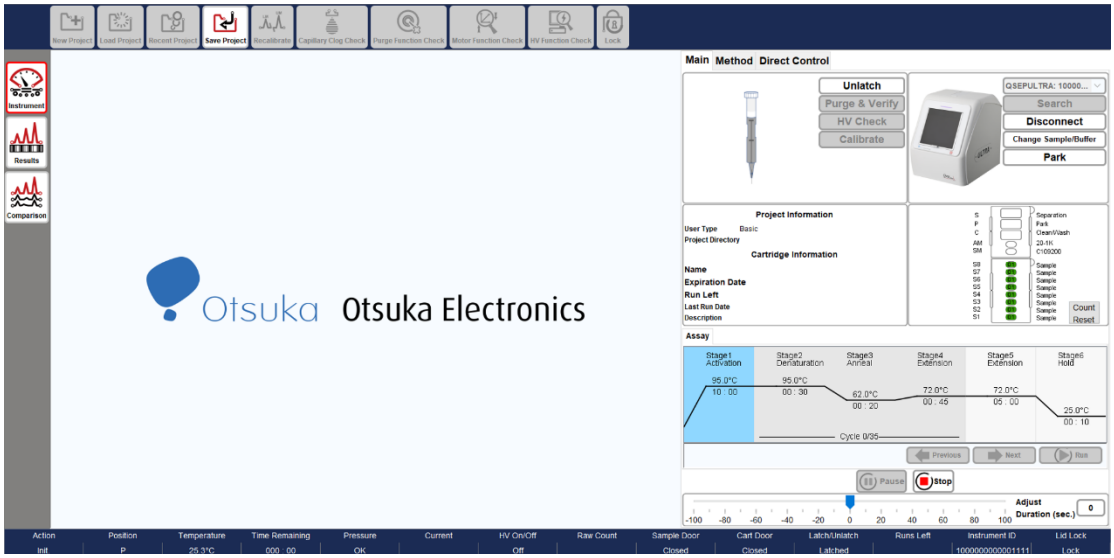
6. Click "**Run**" to start the PCR program and the analysis method.



7. The system will mention **"Please do not open the sample door during PCR!"**. Click **"OK"** to start the PCR.



8. The ongoing stage of the PCR will be shown in blue background.



The results of quality control (NTC and Positive Control)

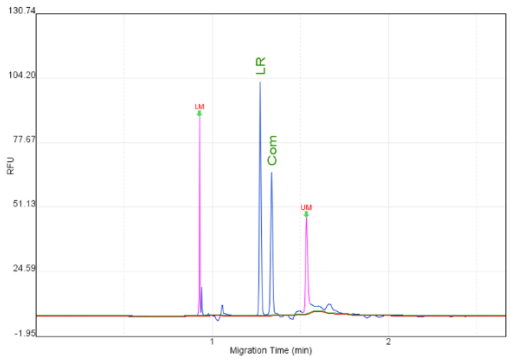
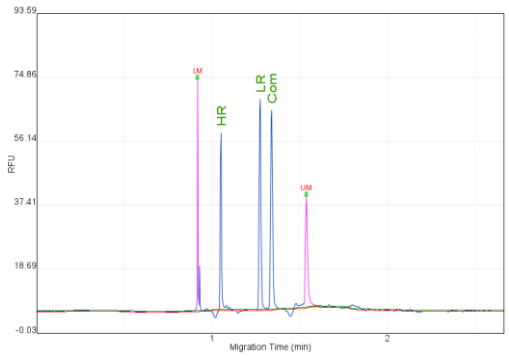
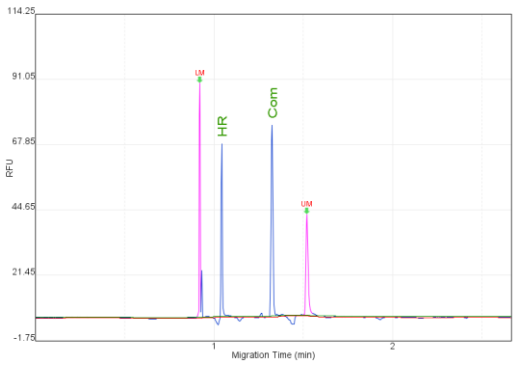
- Ensure that there is no signal of the target sequences revealed in the NTC.
  - If the NTC has any signal, the reagents might be contaminated in the procedure.
- Ensure that all signals of the target sequences are shown in the positive control.
  - If the positive control did not show any signal, it may be caused by the degradation of the reagents. Please contact the distributor in your area.

The result of Direct NQO1 assay

Table 4 The size of the target sequences

NQO1	High Risk (HR)	Low Risk (LR)	Common (Com)
Target size	114 bp	278 bp	338 bp

Table 5 The CE result and interpretation of Direct NQO1 kit assay

Type	Result data
Low Risk	
Medium Risk	
High Risk